TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 USC 371

International Application No.:	PCT/US97/15611
International Filing Date:	September 5, 1997
Priority Date Claimed:	September 6, 1996
Title of Invention:	VITRIFICATION SOLUTIONS FOR SHELF PRESERVATION OF CELLS, TISSUES, ORGANS AND ORGANISMS
Applicant(s) for DO/EO/US:	US

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. (X) This is a **FIRST** submission of items concerning a filing under 35 USC 371.

2. (X) This express request to begin national examination procedures (35 USC 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 USC 371(b) and PCT Articles 22 and 39(1).

3. (X) A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.

4. (X) A copy of the International Application as filed (35 USC 371(c)(2))
   a. () is transmitted herewith (required only if not transmitted by the International Bureau).
   b. () has been transmitted by the International Bureau.
   c. (X) is not required, as the application was filed in the United States Receiving Office (RO/US).

5. (X) Amendments to the claims of the International Application under PCT Article 19 (35 USC 371(c)(3))
   a. () are transmitted herewith (required only if not transmitted by the International Bureau).
   b. () have been transmitted by the International Bureau.
   c. () have not been made; however, the time limit for making such amendments has NOT expired.
   d. (X) have not been made and will not be made.

6. (X) A copy of the International Preliminary Examination Report with any annexes thereto, such as any amendments made under PCT Article 34.

7. (X) International Application as published.

8. (X) Small Entity Statement.

9. (X) PCT request form.

10. (X) International Search Report.

11. (X) A return prepaid postcard.

12. (X) The following fees are submitted:

				FEES
	BASIC FEE			$670
CLAIMS	**NUMBER FILED**	**NUMBER EXTRA**	**RATE**	
Total Claims	24 - 20 =	4 ×	$18	$72
Independent Claims	4 - 3 =	1 ×	$78	$78
Multiple dependent claims(s) (if applicable)			$260	$
	TOTAL OF ABOVE CALCULATIONS		$820	
Reduction by 1/2 for filing by small entity (if applicable). Verified Small Entity statement must also be filed. (NOTE 37 CFR 1.9, 1.27, 1.28)				$410
	TOTAL NATIONAL FEE			$410
	TOTAL FEES ENCLOSED			$410
		amount to be refunded:		$
		amount to be charged:		$

13. (X) A check in the amount of $410 to cover the above fees is enclosed.

14. (X) The Commissioner is hereby authorized to charge only those additional fees which may be required to avoid abandonment of the application, or credit any overpayment to Deposit Account No. 11-1410. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

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Application or Patent No.: Unknown
Filed or Issued: herewith
For: **VITRIFICATION SOLUTIONS FOR SHELF PRESERVATION OF CELLS, TISSUES, ORGANS AND ORGANISMS**

Attorney's Docket No.: UPTINC.015A
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VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL-ENTITY STATUS

I, the undersigned, do hereby declare that:

[X] I am an official of the small business concern empowered to act on behalf of the concern identified below:

NAME OF CONCERN: Universal Preservation Technologies
ADDRESS OF CONCERN: 11045 Roselle St., #C, San Diego, CA 92121

I further declare that the above-identified small business concern qualifies as a small business concern as defined in 13 CFR 121.12, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees to the United States Patent and Trademark Office, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both. I further declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention described in the patent or application identified above.

The individual, concern or organization identified above has not assigned, granted, conveyed or licensed, and is under no obligation under contract or law to assign, grant, convey or license, any rights in the invention to any person who would not qualify as an independent inventor under 37 CFR 1.9(c) if that person had made the invention, or to any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

If the rights held by the above-identified individual, concern or organization are not exclusive, each individual, concern or organization having rights in the invention are identified below. Each such individual, concern or organization must file separate verified statements averring to their status as small entities.

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small-entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b)).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

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SIGNATURE: V. Br DATE: 3/5/99

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VITRIFICATION SOLUTIONS FOR SHELF PRESERVATION OF CELLS, TISSUES, ORGANS AND ORGANISMS

BACKGROUND OF THE INVENTION

1. Field of Invention

This invention relates to the long-term shelf preservation of cells and multicellular specimens by vitrification. The invention is directed to the optimization of vitrification and rehydration solutions, as well as vitrification, and rehydration procedures.

2. Description of the Related Art

Low temperature preservation of cells and multicellular specimens by traditional freezing methods is not uncommon. However, the strong damaging action of ice crystallization limits the effectiveness of such cryogenic methods to the cryopreservation of single cells and multicellular specimens. Vitrification is an alternative approach to cryopreservation that utilizes solidification of samples during cooling, without formation of ice crystals (Fahy, G.M. et al., 1984). Conventionally, cryopreservation by vitrification of single cell (erythocyte, stem cells, sperm, *E. Coli*, yeasts and other cellular microorganisms, etc.) and multicellular specimens provide for storage of cryopreserved samples at -196°C in liquid N_2. However, there is currently a need for reliable methods for long-term shelf preservation at refrigeration or higher temperatures. We believe that development of these methods was not possible because of several generally accepted misconceptions and deficiencies of the prior art that have been addressed by the inventor (Bronshtein, V.L., 1995a).

Effects of dehydration

Ice formation at low temperatures can be avoided only if samples are sufficiently dehydrated. Dehydration is known to damage cells. The damaging effect of dehydration increases with increasing osmotic pressure (concentration) and depends strongly upon whether the vitrification solution contains permeating cryoprotectants. For example, cells normally cannot survive equilibration in

solutions containing only non-permeating solutes in concentration >1 mol/l. However, many types of cells can easily tolerate equilibration in solutions containing permeating cryoprotectants in much higher concentrations. This is because penetration of cryoprotectants protects cells against dehydration damage.

Here, it is important to note that dehydration does not mean a decrease in the cell volume which actually may be very damaging (Meryman, H.T., 1967, Meryman, H.T., 1970). The term "dehydration" means removal of water, or increase in the osmotic pressure. Erroneous use of this term resulted in several misconceptions. For example, as described below, dehydration by itself is not a strong damaging factor. Dehydration may even be a protective factor, as performed according to the present invention.

As shown in Bryant, G. et al. (1992) damage of unloaded specimens during dehydration in vitrification solution is caused by hydration forces occurring between biological macromolecules and membranes when distances between them become small as a result of dehydration. It is believed that loading of cells with permeating cryoprotectants, protects against subsequent dehydration because intracellular cryoprotectant diminishes these forces. Therefore, some amount of intracellular cryoprotectants are required to protect cells during dehydration to high osmotic pressures. For this reason, Rall proposed equilibration of biological specimens in loading solutions of permeating cryoprotectants (dimethylsulfoxide (DMSO), ethylene glycol (EG), propylene glycol (PG), glycerol, etc.) prior to dehydration, in order to reduce the strong damaging effect of dehydration in the vitrification solution (Rall, W.F. et al., 1985a). Unfortunately, the protective effect of loading significantly decreases with increasing time of equilibration in vitrification solution. Currently, this effect is erroneously explained as a direct toxic effect of high concentration of intracellular cryoprotectants.

Apparent toxicity of vitrification solution

Based on the general belief that intracellular cryoprotectants help to vitrify cytosol, and the fact that some intracellular cryoprotectant is required to protect cells during dehydration, penetration of cryoprotectant inside cells may be considered as a beneficial phenomena. A negative aspect of this penetration, considered in the literature, is associated with direct chemical toxicity of cryoprotectants (Fahy et al., 1990). Because the toxicity is believed to be proportional to the concentration of cryoprotectants (not to the amount of cryoprotectants inside a cell) three basic approaches have been proposed to minimize the toxicity (for details see review of Steponkus, P.L. et al., 1992):

1. to use a mixture of different cryoprotectants;

2. to add components that may act as "toxicity neutralizers"; and

3. to identify solutes that will form a glass at a lower concentration.

However, Fahy found that biochemical studies of the toxicity to date have not adequately demonstrated the mechanisms of toxicity (Fahy et al., 1990). This actually means that the direct chemical toxicity of typical permeating cryoprotectants (EG, PG, glycerol and DMSO) is small. Therefore, in agreement with the conclusion of Fahy et al., 1990, present concepts of cryoprotectant toxicity are in need of serious revision.

Recently, Langis, R. et al. (1990) demonstrated that survival of isolated rye protoplast, following a dehydration step, is a function of osmolarity rather than the concentration of vitrification solutions. Based on this observation, Steponkus, P.L. et al. (1992) discussed an alternative strategy for formulating less toxic solutions with lower osmolarity.

As mentioned above, cells can tolerate dehydration in very concentrated vitrification solution for several minutes if they have been loaded with permeating cryoprotectants. However, during long equilibration times in vitrification solution, cell survival decreases with increasing time of equilibration. Because loading of cells with permeating cryoprotectants protects against injury subsequently occurred after dehydration in vitrification solutions, in the case of short dehydration times one may suggest that the injury depends primarily on osmolarity. However, because the concentration of intracellular cryoprotectants that is reached after dehydration increases with increasing osmolarity of vitrification solution, the existing experimental observations do not answer the question whether damage of dehydrated embryos is a result of the increased concentration of intracellular cryoprotectant, or the increase in osmotic pressure. In both cases, however, the questions as to why the injury increases with dehydration time remains to be answered. It is also very important because the time required to complete dehydration of multicellular specimens can be substantially longer than that for individual cells.

Bronshteyn, V.L. et al. (1994) and Steponkus, P.L. et al. (1994) discussed an alterative strategy for formulating less toxic solutions with lower osmolarity. As mentioned above, cells can tolerate dehydration in very concentrated vitrification solution for several minutes if they have been loaded with permeating cryoprotectants. However, during longer equilibration times in vitrification solutions, cell survival decreases with increasing time of equilibration. Because loading of cells with permeating cryoprotectants protects against injury occurring after dehydration in vitrification solution, in the case of short dehydration times, one may suggest that the injury depends primarily on osmolarity. However, because the concentration of intracellular cryoprotectant that is reached after dehydration increases with increasing

osmolarity of vitrification solution, the existing experimental observations do not answer the question of whether damage is a result of the increased concentration of intracellular cryoprotectant or an increase in osmotic pressure. In both cases, no answer is presented as to why injury increases with dehydration time. This answer is very important because the time required to complete dehydration of multicellular specimens can be substantially longer than that for individual cells.

Bronshteyn, V.L. et al. (1994) and Steponkus, P.L. et al. (1994) suggest that a significant part of the apparent toxicity of ethylene glycol-based vitrification for loaded *Drosophila melanogaster* embryos is associated with ethylene glycol permeation (increase in mass of ethylene glycol inside embryos) rather than with chemical toxicity of intra-embryo ethylene glycol, or osmotic pressure of vitrification solution. The injurious effect of permeation of cryoprotectants during equilibration in vitrification solution was also demonstrated in the studies performed with mouse embryos (Zhu, S.E. et al., 1993, Tachikawa, S. et al., 1993 and Kasai, M. et al., 1990). This toxic effect is not related to the increase in intracellular osmotic pressure or biochemical toxicity of cryopreservation because after water efflux from loaded cells, the osmotic pressure and concentration of cryoprotectant inside cells is approximately equal to that outside the cells.

It is believed that influx of penetrating cryoprotectants through the cell membrane during equilibration in vitrification solution containing high concentrations of penetrating cryoprotectants is a main cause of cell damage that occurs during subsequent washing out of the cryoprotectants after cryopreservation.

Kinetics of cryoprotectant permeation inside cells

After the classical work of Kedem, O. et al. (1958) it was generally accepted that the thermodynamic

force responsible for cryoprotectant permeation inside cells is proportional to the cryoprotectant concentration gradient across the cell membrane independent of the composition of the vitrification solution. However, Bronshteyn, V.L. et al. (1994) found that amino acids (glycine and glutamic acid) and carbohydrates (sucrose and sorbitol) significantly diminished ethylene glycol permeation inside *Drosophila melanogaster* embryos. The preventive effect of amino acids was impressive because 1 wt% of glutamic acid + 0.5 wt% glycine practically prevented ethylene glycol permeation inside embryos for up to three hours of equilibration in vitrification solution containing 42 wt% ethylene glycol. The preventive effect of carbohydrates was about four times smaller. These observations show that the approach described in Kedem, O. et al. (1958) and qualitative conclusions obtained based on this model cannot be used to analyze and predict permeation of cryoprotectant inside cells during equilibration in vitrification solution.

Interaction between cryoprotectants and proteins

Timasheff, S.N. (1993) criticized the belief that cryoprotectants form some sort of coating or shell that protects proteins from denaturation during cryopreservation. His criticism was based on the articles of Gekko, K. et al. (1981), Lee, J.C. et al. (1981) and other publications, reporting that cryoprotectants excluded from the surface of proteins. Bronshtein, V.L. (1995b) submitted that the above conclusion of Timasheff and his co-workers is questionable for two reasons. First, the thermodynamic equilibrium in the dialysis experiments of Timasheff and his co-workers cannot be obtained if the hydrostatic pressure inside the dialysis bag is equal to the pressure outside the bag. The suggestion that the effect of this difference in the hydrostatic pressures is negligible is incorrect. Second, amino acids limit penetration of cryoprotectants inside the cell by

decreasing the chemical potential of cryoprotectants in the extracellular aqueous solution (Bronshteyn and Steponkus, 1994). Therefore, cryoprotectant adsorbs at the surface of proteins and partially replaces water molecules hydrating the proteins. The amount of water of hydration, that is, the amount of water at the protein surface that is replaced by molecules of cryoprotectant, increases with increasing concentration of cryoprotectant.

Crowe, J.H. et al. (1990) suggested that freezing and dehydration may be different stress vectors because they found that stabilization of proteins during drying occurs because of an attraction between sugars and proteins. The inventor believes that vitrification of the solution ("shell") at the surface of proteins (and biological membranes) is a general mechanism of protection equally valid for freezing and desiccation.

<u>Effects of intracellular cryoprotectants on the stability of intracellular amorphous state at low temperatures</u>

Steponkus, P.L. et al. (1992) have shown that decreasing osmolarity of the vitrification solution decreases the damaging effect of dehydration in vitrification solution if the dehydration time is several minutes or less. However, to obtain cell survival after cryopreservation, one should successfully vitrify both the extracellular solution and the cytosol. For this reason, Steponkus et al. (1992) suggested that the better cryoprotectant for the loading step is one that allows stable vitrification of cytosol after dehydration in vitrification solution with lower osmolarity. This suggestion was a reflection of a general belief that the presence of cryoprotectants inside cells helps to vitrify cytosol. However, our recent studies (Bronshtein, in preparation) have shown that vitrification temperature of the maximum freeze dehydrated Bovine Serum Aldumin (BSA) solution is T_g=-20°C. In these studies, T_g was estimated as a temperature of detectable onset of ice melting endotherm.

Therefore, T_g in protein solutions is much higher than in solutions of permeating cryoprotectants. This suggests that stability of dehydrated cytoplasm that does not contain cryoprotectants is much higher than that of solutions of permeating cryoprotectants with the same osmotic pressure. This agrees with observations (Steponkus et al., 1992; Langis and Steponkus, 1990) obtained for protoplasts from acclimated rye leaves. They found that the protoplast "loaded with ethylene glycol must be subjected to greater dehydration than those not loaded with ethylene glycol to achieve maximum survival after storage in liquid nitrogen." Bronshteyn and Steponkus (1993) found that intraembryo freezing in non-loaded *Drosophila melanogaster* embryos after dehydration in vitrification solution, occurs at significantly lower temperatures compared to those loaded with 2.125 M ethylene glycol during cooling at 5°C/min. Therefore, contrary to the conventional point of view, addition of low molecular weight cryoprotectants into cytoplasm decreases the stability of the cytoplasm. As such, the present invention is based on scientific theories that are opposite to the prior art described above.

It is, therefore, an object of the present invention to provide a preservation method and a cryoprotectant for cryopreserving cells and multicellular specimens that accounts for the newfound facts that use of low molecular weight cryoprotectants can be detrimental to the cryopreservation process. It is a further object of the present invention to provide a preservation method and a vitrification solution for preserving by vitrification extracellular spaces in the specimen.

SUMMARY OF THE INVENTION

The present invention is directed to a method of preserving cells or multicellular specimens including the step of contacting the specimen with a vitrification solution comprising a permeating cryoprotectant, a non-permeating cryoprotectant and a non-permeating co-solute

that limits the amount of the permeating cryoprotectant that permeates the specimen. The method further includes the step of unloading the specimen by contacting the loaded specimen with a rehydration solution comprising a non-permeating co-solute and, optionally, a permeating cryoprotectant and a non-permeating rehydration cryoprotectant, such that cryoprotectant is removed from the cells of the specimen. Furthermore, the cryoprotectants can be loaded or unloaded in a stepwise manner, in a linear manner, or according to a desired profile.

The present invention is also directed to the vitrification and rehydration solutions for use in connection with the method described above.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is directed to a method for preserving a biological specimen and compositions for achieving the same. Suitable specimens can be single cells (erythocyte, stem cells, sperm, *E. Coli*, yeasts and other cellular microorganisms, etc.) or multicellular tissues (skin, blood vessels, organs, embryos, etc.). The method, vitrification solutions and rehydration solutions described herein minimize toxicity of the vitrification and rehydration solutions and increase intracellular and extracellular vitrification temperatures.

The method includes the step of contacting a specimen or sample with a cryopreservation or vitrification solution. The cryopreservation solution includes a permeating (i.e., low molecular weight) cryoprotectant, a non-permeating (i.e., high molecular weight) cryoprotectant and a non-permeating co-solute that effectively decrease the chemical potential of penetrating cryoprotectants in the vitrification solution. Addition of high molecular weight non-permeating cryoprotectants will increase the vitrification temperature of the cryopreservation solution outside cells. The co-solutes will limit the amount of permeating cryoprotectants that move inside cells and

therefore increase the mass/mass ratio of intracellular protein to permeating cryoprotectant in a dehydrated specimen in cryopreservation solution. This will increase the intracellular vitrification temperature for a given osmotic pressure of cryopreservation solution.

The more co-solutes added, the less cryoprotectant penetrates inside the specimen. The more protein/cryoprotectant ratio inside cells, the higher the intracellular vitrification temperature. However, some minimum amount of cryoprotectant is required inside the cells of the specimen in order to protect the cells against dehydration. For this reason, the concentration of the co-solutes that can be added is limited. The maximum concentration of co-solutes that can be added to cryopreservation solution, to limit penetration of cryoprotectant inside cells, depends upon the minimum amount of cryoprotectant required to protect cells against dehydration in cryopreservation solution. The maximum concentration of co-solutes can be found experimentally for every specific type of permeating cryoprotectants, osmotic pressure of cryopreservation solution, type of co-solute and type of specimen.

As noted above, the invention provides a method for shelf preservation of cells and multicellular specimens at refrigeration or higher temperatures. To increase vitrification temperature outside the cells, cryopreservation solution should contain high molecular weight cryoprotectants, such as dextrans, starches, polyethylene glycol, polyvinylpyrrolidone, Ficol, peptides, etc.

Co-solutes that decrease the chemical potential of penetrating cryoprotectants in aqueous solutions include, but are not limited to:

1. Amino acids: glycine, alanine, glutamic acid, proline, valine, hydroxy-l-proline, beta-aminopropionic acid, aminobutyric acid, beta-ainocaproic acid, aminoisobutyric acid, N-methylglycine, norvaline, and

others that are soluble in water in concentration >0.1 mol/l, and derivatives of amino acids (sarcosine, iminodiacetic acid, hydroxyethyl glycine, etc.) that are soluble in water in concentration >0.1 mol/l.

2. Betaines: betaine and other betaines that are soluble in water in concentration >0.1 mol/l.

3. Carbohydrates: monosaccharide (aldose and ketoses) glyceraldehyde, lyxose, ribose, xylose, galactose, glucose, hexose, mannose, talose, heptose, dihydroxyacetone, pentulose, hexulose, heptulose, octulose, etc., and their derivatives

a. Amino sugars: D-ribose,3-amino-3-deoxy-, chitosamine, fucosamine, etc.;

b. Alditols and inositols: glycerol, erythritol, arabinitol, ribitol, mannitol, iditol, betitol, inositol, etc.;

c. Aldonic, uronic, and aldaric acids that are soluble in water in concentration >0.1 mol/l.; and

d. disaccharides (sucrose, trehalose, etc.).

4. Sugar alcohols (sorbitol, etc.).

To obtain a high intracellular vitrification temperature, the cells should be substantially dehydrated. The dehydration damages the cells due to large repulsive forces between macromolecules inside cells. A small amount of cryoprotectant should be present inside cells in order to decrease these forces. However, the amount of cryoprotectant inside the cells should be kept as low as possible to decrease the toxic effect of the vitrification solution and to increase the intracellular vitrification temperature. All these requirements can be achieved by using cryopreservation solution that contain mixtures of permeating (i.e., low molecular weight) and non-permeating (i.e., high molecular weight) cryoprotectants along with non-permeating co-solutes (amino acids, betaines, sugars, etc. in concentrations from 0.1 - 0.6 mol/l) that effectively decrease the chemical potential of penetrating cryoprotectants in cryopreservation solution.

After dehydration in cryopreservation (vitrification) solution, cells can be stored at a temperature that is lower than the vitrification temperatures both inside and outside the cells of the specimen. Prior to dehydration, cells may be loaded in a low concentration (5-40 wt%), non-damaging solution of permeating cryoprotectant to protect cells from damage during dehydration in cryopreservation solution.

After storage, the samples should be rehydrated and returned to normal physiological medium. In other words, intracellular cryoprotectant should be removed from the cells and exchanged for water. It is believed that damage during rehydration, when cells are transferred from cryopreservation (vitrification) solution to a rehydration (washing) solution, occurs because of an increase in cellular volume beyond initial cellular volumes. To avoid this possibility of damage, one has to include in rehydration solutions, co-solutes, as described above, such as: amino acids, betaines, carbohydrates, or other non-permeating co-solutes that effectively decrease the chemical potential of permeating cryoprotectants in aqueous solutions. The co-solutes are used in concentrations from 0.1 - 0.6 mol/l. Higher co-solute concentrations will more effectively limit the mass of intracellular cryoprotectants, however, when this mass gets very small, the dehydrated cells may be damaged.

The invention allows one to significantly decrease the osmotic pressure of vitrification solution required to obtain a stable vitrification of cells during cooling, to significantly increase extracellular and intracellular vitrification temperatures and the time of cell equilibration (dehydration) in the vitrification solution, without increasing cell damage. This allows one to solve many related problems occurring during equilibration in vitrification solution, storage and rehydration and washing out of intracellular cryoprotectant.

To improve the ability of cells to survive the cryopreservation process described herein, the amounts of permeating cryoprotectant and other components of the cryopreservation solution may be increased in the cryopreservation solution in a stepwise fashion, a linear fashion or according to a desired profile from an initial concentration (≥0%) to an optimal final concentration. The cryopreservation solution and the relative amounts of components thereof may be controlled mechanically or manually. Similarly, to optimize the rehydration process, the contents of the rehydration solution and timing of the rehydration process can be similarly controlled. The optimal initial and final concentrations, as well as the optimum method for increasing the relative concentrations of the components of the cryopreservation and rehydration solutions is determined empirically.

By increasing the intracellular and extracellular vitrification temperatures, one will be able to increase storage temperature up to refrigeration or even room temperature and, therefore, develop method of long-term shelf preservation of cells.

By increasing the equilibration time in vitrification solution, osmotic pressure gradients arising during dehydration of multicellular specimens can be decreased. This is a very important matter because if a portion of cells in the sample is less dehydrated than other portions, it may freeze during subsequent cooling and be damaged.

Limiting the amount of cryoprotectant inside cells simplifies the washing out procedure or completely avoids washing of the intracellular cryoprotectant from cells prior to transfusion or transplantation. This is a very important achievement for blood transfusion, transplantation of embryos and artificial insemination services.

The method of the present invention encompasses dehydration of specimens, cooling samples to a storage

temperature, warming of the samples to ambient temperature, rehydration and washing out of cryoprotectants in rehydration solution, and returning to normal physiological conditions for various medical procedures (transfusions, transplantation, etc.).

The above invention has been described with reference to the preferred embodiment. Obvious modifications and alterations will occur to others upon reading and understanding the preceding detailed description. It is intended that the invention be construed as including all such modifications and alterations insofar as they come within the scope of the appended claims or the equivalents thereof.

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I Claim:

1. A method for preserving a cell or tissue specimen comprising the steps of contacting the specimen with a solution comprising a non-permeating co-solute characterized by its ability to limit the amount of a permeating cryoprotectant to permeate into the specimen.

2. The method for preserving a cell or tissue specimen as claimed in claim 1, wherein the solution further comprises a permeating cryoprotectant and a non-permeating cryoprotectant.

3. The method for preserving a cell or tissue specimen as claimed in claim 1, further comprising the step of contacting the specimen with a cryopreservation solution comprising a permeating cryoprotectant, a non-permeating cryoprotectant and a non-permeating co-solute.

4. The method for preserving a cell or tissue specimen as claimed in claim 2, wherein the cryoprotectant is selected from the group consisting of dimethylsulfoxide, ethylene glycol, propylene glycol and glycerol.

5. The method for preserving a cell or tissue specimen as claimed in claim 2, wherein the non-permeating cryoprotectant is selected from the group consisting of dextrans, starches, polyethylene glycol, polyvinylpyrrolidone, Ficol and peptides.

6. The method for preserving a cell or tissue specimen as claimed in claim 1, wherein the non-permeating co-solute is selected from the group consisting of an amino acid and derivatives thereof, a betaine, a carbohydrate and a sugar alcohol, wherein the carbohydrate is selected from the group consisting of an aldose monosaccharide, a ketose monosaccharide, an amino sugar, an alditol, an inositol,

aidonic, uronic and aldaric acids, disaccharides and polysaccharides.

7. The method for preserving a cell or tissue specimen as claimed in claim 1, wherein the total concentration of non-permeating co-solute in the co-solute solution is between 0.1 and 0.7 mol/l and is equal to a maximum possible concentration that does not substantially damage cells.

8. The method for preserving a cell or tissue specimen as claimed in claim 6, wherein the co-solute is an amino acid.

9. The method for preserving a cell or tissue specimen as claimed in claim 2, wherein the method is performed in two or more stages of contacting the sample with increasingly higher concentrations of the permeating cryoprotectant and the co-solute.

10. The method for preserving a cell or tissue specimen as claimed in claim 2, wherein the method is performed by simultaneously increasing concentrations of both the permeating cryoprotectant and the co-solute from an initial concentration to a final concentration according to a desired profile.

11. The method for preserving a cell or tissue specimen as claimed in claim 2, wherein the rehydration solution further comprises a permeating rehydration cryoprotectant.

12. The method for preserving a cell or tissue specimen as claimed in claim 11, further comprising the step of rehydrating the specimen by contacting the preserved specimen with a rehydration solution comprising a non-permeating rehydration co-solute characterized by its

ability to limit the amount of a permeating cryoprotectant to permeate into the specimen, such that cryoprotectant within the specimen is removed from cells of the specimen.

13. The method for preserving a cell or tissue specimen as claimed in claim 12, wherein the permeating rehydration cryoprotectant is selected from the group consisting of dimethylsulfoxide, ethylene glycol, propylene glycol and glycerol.

14. The method for preserving a cell or tissue specimen as claimed in claim 12, wherein the rehydration step is performed by simultaneously decreasing concentrations of both the permeating rehydration cryoprotectant and the rehydration co-solute from an initial concentration to a final concentration according to a desired profile.

15. The method for preserving a cell or tissue specimen as claimed in claim 12, wherein the non-permeating rehydration co-solute is selected from the group consisting of an amino acid and derivatives thereof, a betaine, a carbohydrate and a sugar alcohol, wherein the carbohydrate is selected from the group consisting of an aldose monosaccharide, a ketose monosaccharide, an amino sugar, an alditol, an inositol, aidonic, uronic and aldaric acids, disaccharides and polysaccharides.

16. The method for preserving a cell or tissue specimen as claimed in claim 1, wherein the contacting step is performed at room temperature or higher.

17. The method for preserving a cell or tissue sample as claimed in claim 1, wherein the specimen can be stably stored at a temperature greater than 4°C.

18. A cryopreservation solution for use in cryopreserving a cell or tissue specimen comprising a permeating cryoprotectant, a non-permeating cryoprotectant and a non-permeating co-solute.

19. The cryopreservation solution as claimed in claim 18, wherein the permeating cryoprotectant is selected from the group consisting of dimethylsulfoxide, ethylene glycol, propylene glycol and glycerol.

20. The cryopreservation solution as claimed in claim 18, wherein the non-permeating cryoprotectant is selected from the group consisting of dextrans, starches, polyethylene glycol, polyvinylpyrrolidone, Ficol and peptides.

21. The cryopreservation solution as claimed in claim 18, wherein the non-permeating co-solute is selected from the group consisting of an amino acid and derivatives thereof a betaine, a carbohydrate and a sugar alcohol, wherein the carbohydrate is selected from the group consisting of an aldose monosaccharide, a ketose monosaccharide, an amino sugar, an alditol, an inositol, aidonic, uronic and aldaric acids, disaccharides and polysaccharides.

22. A rehydration solution for use in rehydrating cryopreserved cell or tissue specimen comprising a permeating rehydration cryoprotectant and a non-permeating rehydration co-solute.

23. The rehydration solution as claimed in claim 22, wherein the permeating rehydration cryoprotectant is selected from the group consisting of dimethylsulfoxide, ethylene glycol, propylene glycol and glycerol.

24. The rehydration solution as claimed in claim 22, wherein the non-permeating rehydration co-solute is selected from the group consisting of an amino acid and derivatives thereof, a betaine, a carbohydrate and a sugar alcohol, wherein the carbohydrate is selected from the group consisting of an aldose monosaccharide, a ketose monosaccharide, an amino sugar, an alditol, an inositol, aidonic, uronic and aldaric acids, disaccharides and polysaccharides.

DECLARATION AND POWER OF ATTORNEY - USA PATENT APPLICATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name;

I believe I am the original, first and sole inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled VITRIFICATION SOLUTIONS FOR SHELF PRESERVATION OF CELLS, TISSUES, ORGANS AND ORGANISMS; the specification of which is attached hereto;

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above;

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56;

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a)-(d) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN APPLICATION(S)			Priority Claimed
No.: **PCT /US97/15611**	Country: **PCT**	Date Filed: **September 5, 1997**	**Yes**

POWER OF ATTORNEY: I hereby appoint the registrants of Knobbe, Martens, Olson & Bear, LLP, 620 Newport Center Drive, Sixteenth Floor, Newport Beach, California 92660, Telephone (949) 760-0404, **Customer No. 20,995**, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith (if this application is assigned, I acknowledge that the appointed individuals do not represent me, and that instead they represent the assignee).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful, false statements may jeopardize the validity of the application or any patent issued thereon.

Page 2 Attorney's Docket No. UPTINC.015A

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